27 April 2020

To: Recipients of M24, 3rd ed.

From: Jennifer K. Adams, MT(ASCP), MSHA
Vice President, Standards and Quality

Subject: Combined Corrections

This notification is intended to inform users of corrections made to CLSI document M24, *Susceptibility Testing of Mycobacteria,* Nocardia *spp., and Other Aerobic Actinomycetes,* 3rd ed. The corrections are described below and shown as highlighted and/or stricken text in the table excerpts.

Correction: 27 April 2020

Appendix F. Stock, Working, and Final Concentrations of Antituberculous Drug Solutions for Agar Proportion:

In the moxifloxacin row, the volume of working concentration to add to 200 mL Middlebrook 7H10 agar is listed incorrectly as "0.5 mL." The volume has been corrected to read "1.0 mL."

Appendix F. Stock, Working, and Final Concentrations of Antituberculous Drug Solutions for Agar Proportion

Antimicrobial Agent	Potency,* µg/mL	Solvent	Stock† Concentration, µg/mL	Working Concentration, µg/mL, From Stock Concentration for Middlebrook 7H10 Agar	Volume of Working Concentration, mL, to Add to 200 mL Middlebrook 7H10 Agar	Final Concentration of Drug in Middlebrook 7H10 Agar
Moxifloxacin	Varies with lot	SDW	10 000	100	~~0.5~~1.0	0.5

Abbreviation: SDW, sterile distilled water.

Clinical and Laboratory Standards Institute
950 West Valley Road, Suite 2500, Wayne, PA 19087 USA | www.clsi.org
P: +1.610.688.0100 Toll Free (US): 877.447.1888 F: +1.610.688.0700

Correction: 28 October 2019

Appendix L. Procedure for Verifying the Inoculum Density for Broth Microdilution Susceptibility Testing of Mycobacteria:

In the step-action table, steps 1 and 2, the volumes are listed incorrectly as "10 µL." The volumes have been corrected to read "1.0 µL."

Step	Action	Comment
1	Using a calibrated ~~10-µL~~1.0-µL loop, plate a single loopful of the final inoculum to be tested onto half of a Middlebrook 7H10 or 7H11 agar plate.	• Label half of the Middlebrook 7H10 or 7H11 agar plate as 1× (undiluted) and the other half as 1:50 (diluted). • Vortex the final inoculum to be tested before removing the ~~10-µL~~1.0-µL aliquot. • Plate the entire contents of the loop.
2	Using the same calibrated ~~10-µL~~1.0-µL loop, dilute ~~10 µL~~1.0 µL of the final inoculum 1:50 in sterile water and inoculate to the other half of the Middlebrook 7H10 or 7H11 agar plate used in step 1.	• Vortex the final inoculum to be tested before removing the ~~10 µL~~1.0-µL aliquot and also the 1:50 dilution before plating. • Plate the entire contents of the loop.

In Table L1, the volume in the first and second column headings is listed incorrectly as "10 µL." The volume has been corrected to read "1.0 µL."

Table L1. Interpreting Results of the Inoculum Verification Procedure

Number of Colonies on Plate, ~~10 µL,~~ 1.0 µL, 1×	Number of Colonies on Plate, ~~10 µL,~~ 1.0 µL, 1:50	Estimated CFU/mL	Interpretation
< 50	0	$< 5 \times 10^4$	Inoculum too low; repeat test
50–100	0–2	5×10^4 – 1×10^5	Acceptable
> 100	≤ 10	1×10^5 – 5×10^5	Acceptable
> 100	> 10	5×10^5 – 1×10^6	Acceptable
> 100	> 100	$> 10^6$	Inoculum too high; repeat test

Abbreviation: CFU, colony-forming unit(s).

If you require any additional clarification regarding these corrections, please contact CLSI Customer Service (customerservice@clsi.org).

We appreciate your commitment to CLSI and regret any inconvenience.